

Docteur Fabrice NESSLANY Laboratoire de Toxicologie Génétique

Tél.: 33 (0)3 20 87 72 72 Fax: 33 (0)3 20 87 73 10

e-mail: fabrice.nesslany@pasteur-lille.fr

### **Final Study Report**

#### CONFIDENTIAL

In Vivo Erythrocytes-BASED Pig-A GENE MUTATION ASSAY
(Performed in Mouse Somatic Cells - Two sampling times)

Combined to the
In Vivo MAMMALIAN ALKALINE COMET ASSAY
(Performed in Mouse Circulating Blood Cells - One sampling time)

(Five treatments followed by 3 co-treatments)

Study Number FSR-IPL 160901

**Study Completion** 17 February 2017

Test Item
ADN Telomeractives®

Study Director Dr. Sophie SIMAR

> Sponsor HBN

## TEST FACILITY INSTITUT PASTEUR DE LILLE

Genetic Toxicology Laboratory 1, rue du Professeur Calmette - BP. 245 59019 LILLE CEDEX

SPONSOR HBN

1 rue des pénitents blancs 31010 TOULOUSE SPONSOR REPRESENTATIVE Mr. Gérald GOUDET

#### TABLE OF CONTENTS

STUDY INFORMATION	
GOOD LABORATORY PRACTICE COMPLIANCE STATEMENT AND REPORT AUTHENT	ΓΙCATION4
QUALITY ASSURANCE STATEMENT	6
ARCHIVE STATEMENT	8
SUMMARY	9
1 PURPOSE OF THE STUDY	14
2 PRINCIPLE	
2.1 In vivo Erythrocytes-Based Pig-A Gene mutation assay	
2.2 In vivo comet assay	
3 ANIMALS AND HUSBANDRY	
3.1 Reason for the choice of the reactive system	
3.2 Animals	
3.3 Husbandry	
4 TEST ITEM AND VEHICLE INFORMATION	15
4.1 Test item	
4.2 Vehicle	
4.3 Formulation of the test item	
5 DETERMINATION OF TEST ITEM CONCENTRATION IN DOSING FORMULATIONS	
6 PRELIMINARY TOXICITY TEST	
7 TREATMENTS FOR THE DEFINITIVE GENOTOXICITY ASSAYS	
8 IN VIVO ERYTHROCYTES-BASED PIG-A GENE MUTATION ASSAY	
8.1 Sample preparation	
8.2 Instrument calibration standards	
8.3 Data acquisition	
8.4 Expression of the results and statistical analysis	
8.5 Acceptance criteria for the results	
8.6 Interpretation of the results	
8.7 Results for the <i>in vivo</i> erythrocytes based Pig-A gene mutation assay	
9 COMET ASSAY	
9.1 Cell sampling	
9.2 Protocol for the Comet assay	
9.3 Image analysis	
9.4 Tail parameters	
9.5 Determination of the cytotoxicity of the test item	
9.6 Expression of the results and statistical analysis	
9.7 Acceptance criteria for the results	
9.8 Interpretation of the results	24
9.9 Results for the Comet Assay	24
10 DETERMINATION OF SYSTEMIC EXPOSURE	25
11 GENERAL COMMENTS	25
12 STUDY PLAN ADHERENCE	25
12.1 Deviations	25
12.2 Notes	26
13 CONCLUSION	27
14 REFERENCES	28
Appendix No. 1: Recapitulative Tables	
Appendix No. 2: Individual results for animal weight	
Appendix No. 3a: Individual results for the Pig-A assay	
Appendix No. 3b: Statistical analysis for the Pig-A assay	
Appendix No. 3c: Historical data for the Pig-A assay	
Appendix No. 4a: Individual data for the comet assay	
Appendix No. 4b: individual data for percentage of DNA in tail	
Appendix No. 4c: Coding slide	
Appendix No. 4d: Non-parametric statistical analysis	
Appendix No. 5: Certificate of analysis	
Appendix No. 6: Final Study Plan FSP-IPL 160901	78
Appendix No. 7: Amendment No. 1 to Final Study Plan FSP-IPL 160901	103

## GOOD LABORATORY PRACTICE COMPLIANCE STATEMENT AND REPORT AUTHENTICATION

The work described in this report was performed according to the agreed study plan and with the Standard Operating Procedures of the test facility, unless otherwise stated, and was conducted in accordance with:

- OECD Principles of Good Laboratory Practice (as revised in 1997), ENV/MC/CHEM(98)17;
- GLP departmental order 14/3/2000 (Official Journal of 23<sup>rd</sup> March 2000);
- EC Commission Directive 2004/10/EC of 11<sup>th</sup> February 2004 (Official Journal No. L050);
- Application of the OECD Principles of GLP to Computerised Systems, No. 10 Consensus Document of the Working Group on Good Laboratory Practice, OECD/GD(95)115.

I consider the data generated and reported to be valid and I declare that this report is a true and accurate record of the results obtained.

As described in the Study Plan, the sponsor certifies that the test item to be tested provided by HBN is identical to the test item described in the Final Study Plan.

Note: No Analytical Certificate of the test item was provided.

No data about Composition (or concentration) or Stability in storage conditions was provided. This constitutes a deviation to the Good Laboratory Practices (OECD, 1997: § 6.2, Characterisation).

No control of concentration in dosing formulation was performed. This also constitutes a deviation to the recommendations of the Good Laboratory Practices (OECD, 1997: §6.2, Characterisation).

Nevertheless, taking into account the nature of the test item (i.e. plant extract), this deviation was considered only as a minor deviation.

The study was performed at the Toxicology Department of Institut Pasteur de Lille for genotoxicity assays.

The computer applications used to acquire and derive data for this study included Excel® and Comet assay IV. These applications have been validated in the laboratory (Conformity certificates F-TOX-INF-025 and 024).

Otherwise, the computer application used to calculate mutation frequencies and percent RET was provided by the Manufacturer (Litron Laboratories Ltd) of the *In Vivo* MutaFlow Kits (*i.e.* kit used for the Pig-A test). This application was not validated in the laboratory.

Submitted by:

Study director

Dr. Sophie SIMAR

Agreement of the establishment for realizing experiments on living vertebrate animals No. B 59-350009

STUDY

In Vivo ERYTHROCYTES-BASED Pig-A GENE MUTATION ASSAY

(Performed in Mouse somatic cells - Two sampling times)

Combined to the

In Vivo MAMMALIAN ALKALINE COMET ASSAY

(Performed in Mouse Circulating blood cells - One sampling time)

(Five treatments followed by 3 co-treatments)

**TEST ITEM** 

**ADN Telomeractives®** 

SPONSOR

**HBN** 

This report was reviewed and approved by:

Test Facility Management

Dr. Fabrice NESSLANY

Head of Toxicology Department

Date

Signature

Deputy Study Director

Mrs. Gwendoline MORDACQ

# In Vivo ERYTHROCYTES-BASED Pig-A GENE MUTATION ASSAY (Performed in Mouse somatic cells - Two sampling times) Combined to the

#### In Vivo MAMMALIAN ALKALINE COMET ASSAY

(Performed in Mouse Circulating blood cells - One sampling time)
(Five treatments followed by 3 co-treatments)

#### **SUMMARY**

SPONSOR : HBN

TEST ITEM : ADN Telomeractives®

BATCH NUMBER : N002

STUDY LOCATION : INSTITUT PASTEUR DE LILLE

Genetic Toxicology Laboratory

1. rue du Professeur Calmette - B.P. 245

59019 LILLE CEDEX FRANCE

#### THIS STUDY WAS CARRIED OUT IN COMPLIANCE WITH GOOD LABORATORY PRACTICE REGULATIONS

Study initiation date (date Study Director signed Study Plan): 27/09/2016

Main assay

 Treatments (with the test item alone):
 17 to 21/10/2016

 Co-treatments (test item +/- ENU)
 24 to 26/10/2016

 Sampling for Comet assay
 26/10/2016

 D31/32 sampling for Pig-A
 23&24/11/2016

 D44 sampling for Pig-A
 07/12/2016

 Experimental completion date:
 21/12/2016

 Study completion
 17/02/2017

#### AIM

The evaluation of the protective potential, eg. fight against primary DNA damage and/or optimization of DNA repair capability, of the ADN Telomeractives® test sample was studied using two different endpoints: the measurement of mutant frequency by *in vivo* Erythrocytes-Based Pig-A Gene mutation assay and the evaluation of primary DNA damage by the *in vivo* Comet assay following the alkaline version (pH > 13) in circulating blood cells in mice treated with both the test item and the positive reference substance ethylnitrosourea (ENU).

#### **CONCLUSION**

The test item ADN Telomeractives® (batch N002), provided by HBN, was investigated for its protective potential against DNA damaging agent, eg. fight against primary DNA damage and/or optimization of DNA repair capability, by the means of the evaluation of primary DNA damage by in vivo Comet assay following the alkaline version (pH > 13) in circulating blood cells based on OECD Guideline (No. 489, 2014) and the in vivo Erythrocytes-Based Pig-A Gene mutation assay, in male OF1 mice.

Animals were pre-treated with the test item alone at dose levels of 550 and 55 mg/kg. Oral treatments were carried out once a day for 5 consecutive days, 24 hours apart. Then, after 2 days without any treatment, mice were treated thrice, 24-hours apart, with the test item at the 2 same dose levels. One hour after each treatment with the test item, animals were treated with either the DNA damaging agent ethylnitrosourea or its vehicle.

The validity criteria for the results were fulfilled. The study was thus considered as valid.

Under our experimental conditions, ADN Telomeractives® induced no mutagenic activity in circulating blood cells from OF1 male mice. Furthermore, the test item did not present DNA strand breaks and/or alkali-labile sites inducer activities toward the circulating blood cells from male OF1 mice,

On the other hand, under these operating conditions, in vivo, ADN Telomeractives® decreased both DNA fragmentation and mutation frequency induced by ethylnitrosourea, a well-known potent mutagen/carcinogen. Therefore, ADN Telomeractives® is considered to have a protectant potential against primary DNA damage and mutation induced by a strong mutagenic substance ENU.

Figure 1

In Vivo ERYTHROCYTES-BASED Pig-A GENE MUTATION ASSAY (Performed in Mouse somatic cells - Two sampling times)

Combined to the

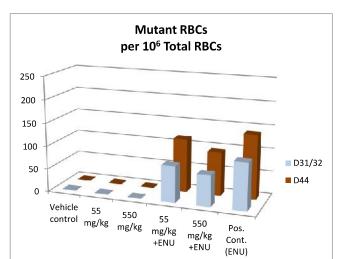
#### In Vivo MAMMALIAN ALKALINE COMET ASSAY

(Performed in Mouse Circulating blood cells - One sampling time)
(Five treatments followed by 3 co-treatments)

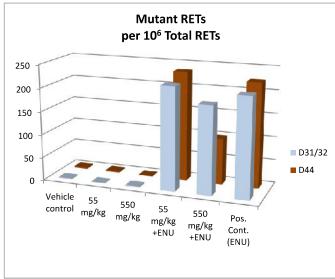
Sponsor: HBN

Test item: ADN Telomeractives®

Vehicle: CMC at 0.5% in sterile water



Species: Mouse
Strain: OF1
Route: oral
Volume: 10 mL/kg\*



<sup>\*</sup> Phase I: 10 mL/kg/day (x5) - Phase II: 10 mL/kg/day (test item or vehicle control) + 10 mL/kg/day (x3) (ENU or sterile water)